

## Effects of chronic administration of 7-benzylidene-7-dehydronaltrexone and naltriben on the antinociceptive actions of $\delta_1$ - and $\delta_2$ -opioid receptor agonists

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### Abstract

The effects of chronic administration of 7-benzylidene-7-dehydronaltrexone, a  $\delta_1$ -opioid receptor antagonist and naltriben, a  $\delta_2$ -opioid receptor antagonist, on the antinociceptive responses to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II,  $\delta_1$ - and  $\delta_2$ -opioid receptor agonists, respectively, were determined in the mouse. Female B6C3F1 mice were given 7-benzylidene-7-dehydronaltrexone (3 mg/kg/day), naltriben (1 mg/kg/day) or the vehicle by subcutaneously implanted Alzet osmotic minipumps for 7 days. Both [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II administered intracerebroventricularly (i.c.v.) produced antinociceptive as measured by the tail-flick test with ED<sub>50</sub> values of 6.76 and 6.68  $\mu$ g/mouse, respectively. Chronic administration of 7-benzylidene-7-dehydronaltrexone lowered the ED<sub>50</sub> of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin but not of [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II. Chronic administration of naltriben lowered the ED<sub>50</sub> of [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II but had no effect on the ED<sub>50</sub> of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin. The binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to whole brain membranes of chronic 7-benzylidene-7-dehydronaltrexone-treated mice did not differ from chronic vehicle-treated mice. On the other hand, chronic administration of naltriben resulted in slight but reproducible elevation in the  $B_{max}$  value of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to bind to whole brain membranes in comparison to vehicle-injected controls. The results suggest that chronic treatment with  $\delta_1$ - and  $\delta_2$ -opioid receptor antagonist cause behavioral supersensitivity to their agonists, respectively, and provides further evidence for the existence of  $\delta$ -opioid receptor subtypes.

**Keywords:**  $\delta$ -Opioid receptor, agonist, antagonist;  $\delta_1$ -Opioid receptor;  $\delta_2$ -Opioid receptor; Antinociception; [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]Enkephalin

### 1. Introduction

Opioid drugs produce their pharmacological actions by interacting with three types of receptors, viz.  $\mu$ ,  $\delta$  and  $\kappa$  (Bhargava, 1994). The possible existence of subtypes of these receptors has been suggested. Thus,  $\mu$ - and  $\kappa$ -opioid receptors have been further classified into  $\mu_1$  and  $\mu_2$ , and  $\kappa_1$ ,  $\kappa_2$  and  $\kappa_3$  based on behavioral and biochemical differences in the activity of related compounds. Recently, two subtypes of  $\delta$ -opioid receptors have been identified, and labeled as  $\delta_1$  and  $\delta_2$ . The prototypical agonists at these receptors are [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II, respectively (Mattia et al., 1991). Such classification is based on the fact that naltriben, a highly selective  $\delta_2$ -opioid receptor antagonist, is more effective in

antagonizing the antinociceptive action of [D-Ser<sup>2</sup>, Leu<sup>5</sup>, Thr<sup>6</sup>]enkephalin than that of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in mice (Sofuoglu et al., 1991). Lack of cross-tolerance between [D-Ser<sup>2</sup>, Leu<sup>5</sup>, Thr<sup>6</sup>]enkephalin and [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin or between [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II provided additional evidence for  $\delta$ -opioid receptor heterogeneity (Mattia et al., 1991). Differential antagonism of  $\delta$ -opioid receptor agonists of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II-induced antinociceptive by the irreversible  $\delta$ -opioid receptor antagonists, [D-Ala<sup>2</sup>, Leu<sup>5</sup>, Cys<sup>6</sup>]enkephalin and naltrindole-5<sup>1</sup>-isothiocyanate also indicated the possibility of  $\delta$ -opioid receptor subtypes (Jiang et al., 1991).

Recent studies in our laboratory have provided immunological evidence for the existence of subtypes of  $\delta$ -opioid receptors. In general, [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin, a  $\delta_1$ -opioid receptor agonist was found to have greater im-

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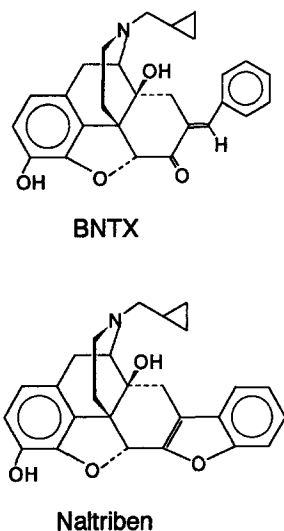


Fig. 1. Chemical structures of 7-benzylidene-7-dehydronaltrexone (BNTX) and naltriben.

munostimulant activity than [D-Ser<sup>2</sup>, Leu<sup>5</sup>, Thr<sup>6</sup>]enkephalin or other  $\delta_2$ -opioid receptor agonists (Bhargava et al., 1995a). On the other hand, 7-benzylidene-7-dehydronaltrexone, a highly selective  $\delta_1$ -opioid receptor antagonist produced immunosuppression whereas naltriben was devoid of any effect on cellular immune function (House et al., 1995).

Since 7-benzylidene-7-dehydronaltrexone and naltriben have been designated as highly selective  $\delta_1$ - and  $\delta_2$ -opioid receptor antagonists, respectively, we tested the hypothesis that chronic blockade of  $\delta_1$ - and  $\delta_2$ -opioid receptors should up-regulate their own receptors. For this purpose, the effects of chronic administration of 7-benzylidene-7-dehydronaltrexone and naltriben (Fig. 1), by using osmotic minipumps, on the antinociceptive responses to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II and the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to brain membranes were determined in mice.

## 2. Materials and methods

### 2.1. Animals

Female, 6–8-week-old B<sub>6</sub>C<sub>3</sub>F<sub>1</sub> (B57BL6XC3H) mice obtained from Charles River Laboratories (Wilmington, MA, USA) were acclimated to a room with controlled ambient temperature (23 ± 1°C), humidity (50 ± 10%) and a 12-h dark-light cycle (light 06:00–18:00 h). The animals were housed under these conditions for at least 4 days before being used. The animals were given food and water ad libitum.

### 2.2. Drugs

7-Benzylidene-7-dehydronaltrexone HCl, naltriben methane sulfonate, [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>,

Glu<sup>4</sup>]deltorphin II were supplied by the Research Technology Branch, National Institute on Drug Abuse (Rockville, MD, USA) through the courtesy of Mr. Kevin Gormley. 7-Benzylidene-7-dehydronaltrexone and naltriben were dissolved in 10% dimethylsulfoxide in saline and administered subcutaneously to mice via Alzet osmotic minipumps (Model 1007 D) which delivered the drug solution at a rate of 0.5  $\mu$ l/h for 7 days. The daily dose of 7-benzylidene-7-dehydronaltrexone and naltriben were 3 and 1 mg/kg, respectively. Mice which served as controls were implanted with osmotic minipumps filled with the vehicle. The osmotic minipumps were removed 7 days after their implantation. The implantation and removal of the pumps were done under light ether anesthesia.

[D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II were dissolved in water and 10% dimethylsulfoxide in water solution, respectively, and were injected i.c.v. under light ether anesthesia in a volume of 5  $\mu$ l/mouse according to the method of Haley and McCormick (1957) as described previously (Bhargava and Zhao, 1996; Zhao and Bhargava, 1996). The coordinates for the injection site were 2 mm lateral and caudal to bregma at a depth of 3 mm by using a 10- $\mu$ l Hamilton syringe with a 27-gauge needle.

### 2.3. Measurement of the antinociceptive response

The antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II was measured by the tail-flick test as described previously (D'Amour and Smith, 1941; Bhargava and Thorat, 1994; Bhargava et al., 1995b). At the beginning of the study, the light intensity in the tail-flick apparatus was adjusted such that the mean basal latencies for the tail-flick response were approximately 2 s. To minimize tail skin tissue damage, the cut-off time was set at 10 s. The tail-flick latencies were determined before and 15 min after the i.c.v. injection of an appropriate dose of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin or [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II. A delay of 3 times the tail-flick latency over the basal value after i.c.v. injection of the drug was considered to be a positive response. Percentage of mice exhibiting antinociceptive response was calculated for each dose of the drug. Ten mice were used for each dose of the drug. Dose-response curves were constructed and the ED<sub>50</sub> values and their confidence limits were determined by the method of Litchfield and Wilcoxon (1949).

### 2.4. Effects of chronic administration of 7-benzylidene-7-dehydronaltrexone and naltriben on the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in mice

Seven days after the continuous administration of 7-benzylidene-7-dehydronaltrexone, naltriben or the vehicle, the osmotic minipumps were removed under light ether

anesthesia. The antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II were determined 6 h after the removal of the osmotic minipumps. The ED<sub>50</sub> values and their 95% confidence limits of the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in mice injected with the vehicle, 7-benzylidene-7-dehydronaltrexone or naltriben were determined as described above.

## 2.5. Determination of binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to brain membranes

### 2.5.1. Membrane preparation

Mice were sacrificed and the brain quickly excised into an ice-cold Petri dish. The cerebellum was removed and the remainder of the brain was homogenized in 60 vols. of ice-cold Tris-HCl buffer (0.05 M, pH 7.4) using a Brinkman polytron homogenizer (setting 5 for 20 s). The homogenate was centrifuged at 49 000 × *g* for 15 min in a refrigerated Sorvall RC-5B centrifuge. The process was repeated once more. After the second centrifugation, the pellet was stored at –80°C. For the binding assay, the pellet was suspended in 25 vols. of Tris-HCl buffer by homogenizing for 15 s as described above.

### 2.5.2. Binding assays

The binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin was performed as described previously (Bhargava et al., 1991; Magnan et al., 1982). Binding was carried out in a total volume of 0.5 ml which contained 0.2 ml of homogenate (450–500 µg protein) and 0.05 M Tris-HCl buffer. In saturation experiments, the [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin concentration range was 1.0–40.0 nM. All binding assays were done in duplicate at 37°C for 60 min. Binding was terminated by rapidly filtering the contents of the incubation tubes through Whatman GF/B glass fiber filter under reduced pressure using a Brandell cell harvester (model M-24R). The filters were washed twice with 5 ml of the ice-cold 0.05 M Tris-HCl buffer. The filters were transferred to liquid scintillation vials containing 5 ml of SCINT-AXF scintillation fluid (Packard Instruments, Meriden, CT, USA). After an overnight equilibration period, the radioactivity in the samples was determined using a Packard liquid scintillation counter (model 4640) with a 54% counting efficiency. Specific binding was defined as the difference in binding observed in the absence and presence of 3.5 µM unlabeled [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin. The concentration of protein in the samples was determined by employing the method of Lowry et al. (1951).

Receptor density (*B*<sub>max</sub>) and apparent dissociation constant (*K*<sub>d</sub>) for the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to brain membranes were determined from the saturation curves and Scatchard plots using the LIGAND program (Munson and Rodbard, 1980). The results were expressed as mean ± S.E.M. Six to 8 mice were used to determine the binding constants.

### 2.5.3. Effect of chronic administration of 7-benzylidene-7-dehydronaltrexone and naltriben on the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to mouse brain membranes

Seven days after the continuous administration on 7-benzylidene-7-dehydronaltrexone and naltriben or the vehicle, the osmotic minipumps were removed under light ether anesthesia. Six hours later, mice were decapitated and the brains quickly removed for the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin as described above. The *B*<sub>max</sub> and *K*<sub>d</sub> values were determined in brains from all the treatment groups. The differences in the binding constants of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in vehicle- and drug-treated mice was determined by Student's *t*-test. A value of *P* < 0.05 was considered to be statistically significant.

## 3. Results

### 3.1. Effect of chronic administration of 7-benzylidene-7-dehydronaltrexone and naltriben on the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin

Chronic administration of 7-benzylidene-7-dehydronaltrexone enhanced the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin as evidenced by a leftward shift in its dose-response curve in 7-benzylidene-7-dehydronaltrexone-treated mice in comparison to vehicle-injected mice. The ED<sub>50</sub> value of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in vehicle-injected mice was 6.76 µg/mouse which was decreased by almost 50% to 3.41 µg/mouse in 7-benzylidene-7-dehydronaltrexone-treated mice (Table 1). On the other hand, the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin was not altered in mice treated chronically with naltriben (ED<sub>50</sub> value 6.82 µg/mouse) when compared to vehicle-injected mice. The dose-response curves for [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in vehicle- and naltriben-treated mice did not differ and thus the ED<sub>50</sub> values of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin also did not change (Table 1).

Table 1

Effect of chronic administration of 7-benzylidene-7-dehydronaltrexone (BNTX) and naltriben on the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in B<sub>6</sub>C<sub>3</sub>F<sub>1</sub> mice

Treatment	ED <sub>50</sub> of [D-Pen <sup>2</sup> , D-Pen <sup>5</sup> ]enkephalin (µg/mouse)
Vehicle	6.76 (4.12–11.06) <sup>a</sup>
BNTX	3.41 (2.03–5.73) <sup>a,b</sup>
Naltriben	6.82 (4.05–11.47) <sup>a</sup>

Mice were administered vehicle, 7-benzylidene-7-dehydronaltrexone or naltriben by Alzet osmotic minipumps for 7 days as described in the text. The antinociceptive response to different doses of i.c.v. injected [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II was determined by the tail-flick test.

<sup>a</sup> 95% confidence limits.

<sup>b</sup> *P* < 0.05 vs. vehicle-treated group.

Table 2

Effect of chronic administration of 7-benzylidene-7-dehydronaltrexone (BNTX) and naltriben on the antinociceptive response to [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in B<sub>6</sub>C<sub>3</sub>F<sub>1</sub> mice

Treatment	ED <sub>50</sub> of [D-Ala <sup>2</sup> , Glu <sup>4</sup> ]deltorphin II (μg/mouse)
Vehicle	6.68 (5.18–8.62) <sup>a</sup>
BNTX	6.67 (4.99–8.92) <sup>a</sup>
Naltriben	3.72 (2.48–5.58) <sup>a,b</sup>

Mice were administered vehicle, 7-benzylidene-7-dehydronaltrexone or naltriben by Alzet osmotic minipumps for 7 days as described in the text. The antinociceptive response to different doses of i.c.v. injected [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II was determined by the tail-flick test.

<sup>a</sup> 95% confidence limits.

<sup>b</sup> *P* < 0.05 vs. vehicle-injected controls.

### 3.2. Effect of chronic administration of 7-benzylidene-7-dehydronaltrexone and naltriben on the antinociceptive response to [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II

Chronic administration of naltriben enhanced the antinociceptive response to [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II as evidenced by a leftward shift in its dose-response curve in naltriben-treated mice in comparison to vehicle-injected mice. The ED<sub>50</sub> value of [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in naltriben-treated mice was 3.72 μg/mouse, which was significantly lower than in mice treated chronically with vehicle (6.68 μg/mouse) (Table 2). The antinociceptive response to [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II, on the other hand, was not affected in mice treated chronically with 7-benzylidene-7-dehydronaltrexone (6.67 μg/mouse) when compared to vehicle-injected mice. The dose-response curves for [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in vehicle and 7-benzylidene-7-dehydronaltrexone-treated overlapped; consequently the ED<sub>50</sub> values for [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in the two treatment groups were virtually identical (Table 2).

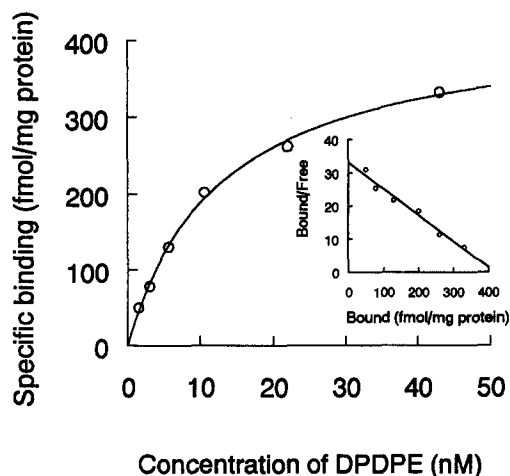


Fig. 2. A typical saturation curve and Scatchard plot (inset) for the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to mouse brain membranes.

Table 3

Effect of chronic administration of 7-benzylidene-7-dehydronaltrexone (BNTX) and naltriben on the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to brain membranes of B<sub>6</sub>C<sub>3</sub>F<sub>1</sub> mice

Treatment	[ <sup>3</sup> H][D-Pen <sup>2</sup> , D-Pen <sup>5</sup> ]enkephalin binding constants (mean ± S.E.M. <i>n</i> = 6–8)	
	<i>B</i> <sub>max</sub> (fmol/mg protein)	<i>K</i> <sub>d</sub> (nM)
Vehicle	371 ± 13	6.74 ± 0.30
BNTX	379 ± 17	6.80 ± 0.60
Naltriben	412 ± 10 *	7.49 ± 0.68

The binding [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin was carried out in a concentration of 1.0–40.0 nM. Specific binding was defined as the binding observed in the absence and presence of 3.5 μM unlabeled [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin.

\* *P* < 0.05 vs. vehicle-injected controls.

### 3.3. Effect of chronic administration on 7-benzylidene-7-dehydronaltrexone and naltriben on the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to mouse brain membranes

[<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin bound to the mouse brain membranes at a single high-affinity site with a *B*<sub>max</sub> value of 371 fmol/mg protein and a *K*<sub>d</sub> value of 6.7 nM. A typical Scatchard plot of the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to brain membranes is shown in Fig. 2. As can be seen in Table 3, chronic administration of 7-benzylidene-7-dehydronaltrexone did not alter the binding constants of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in the mouse brain. However, a small (10%) but significant increase in the *B*<sub>max</sub> value of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin was observed in mice treated chronically with naltriben; *K*<sub>d</sub> value was unaffected.

## 4. Discussion

The present studies clearly indicate that the chronic blockade of δ<sub>1</sub>-opioid receptor by 7-benzylidene-7-dehydronaltrexone enhances the antinociceptive action of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin, a δ<sub>1</sub>-opioid receptor agonist. However, such a treatment had no effect on the antinociceptive response to [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II, a highly selective δ<sub>2</sub>-opioid receptor agonist. On the other hand, chronic blockade of the δ<sub>2</sub>-opioid receptor by naltriben enhanced the antinociceptive action of [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II, but had no effect on the antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin. These results provide evidence for the functional up-regulation of δ<sub>1</sub>- and δ<sub>2</sub>-opioid receptors as a result of their chronic blockade.

The results of the present studies further confirm the existence of δ-opioid receptors subtypes. Previous studies have also provided behavioral evidences for the existence of subtypes of δ-opioid receptors. Thus, naltriben was shown to antagonize the antinociceptive action of [D-Ser<sup>2</sup>, Leu<sup>5</sup>, Thr<sup>6</sup>]enkephalin but not of DADLE and [D-Pen<sup>2</sup>,

D-Pen<sup>5</sup>]enkephalin (Sofuoglu et al., 1991). Additionally, a lack of cross-tolerance between [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II has been demonstrated (Mattia et al., 1991). However, to date, little biochemical (binding) evidence has been provided for such a phenomenon.

To determine whether the enhanced responses to  $\delta_1$ - and  $\delta_2$ -opioid receptor agonists in 7-benzylidene-7-dehydronaltrexone- and naltriben-treated mice, respectively, were related to alterations in the  $\delta$ -opioid receptor characteristics, the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin to brain membranes was determined. It was clear from the present studies that  $B_{\max}$  and  $K_d$  values of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin were unaffected by 7-benzylidene-7-dehydronaltrexone treatment. However, there was a 10% increase in  $B_{\max}$  of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in the brain of naltriben-treated mice over the controls but although the  $K_d$  values did not differ. This effect, although small was consistently reproducible. Thus, the enhanced antinociceptive response to [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in mice treated chronically with 7-benzylidene-7-dehydronaltrexone can not be explained by the changes in the characteristics of  $\delta$ -opioid receptors labeled with [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin. Similarly, a 45% decrease in the ED<sub>50</sub> value of [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II could not be accounted for by a 10% increase in the  $B_{\max}$  value of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin in naltriben-treated mice. Since there is no cross-tolerance between [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II, a 10% increase in the  $B_{\max}$  of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin would not account for the enhanced response to [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II in naltriben-treated mice.

Although naltriben could distinguish between [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [D-Ser<sup>2</sup>, Leu<sup>5</sup>, Thr<sup>6</sup>]enkephalin behaviorally, it was shown to compete equally well against the binding of [<sup>3</sup>H][D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin and [<sup>3</sup>H][D-Ser<sup>2</sup>, Leu<sup>5</sup>, Thr<sup>6</sup>]enkephalin in brain membranes (Sofuoglu et al., 1992). Thus, binding studies have not been able to distinguish between  $\delta_1$ - and  $\delta_2$ -opioid receptors.

In another study, chronic administration of naltrexone, a non-specific opioid receptor antagonist, was shown to shift the antinociceptive dose-response curve for [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II to the left, i.e. there was supersensitivity to the antinociceptive effect of [D-Ala<sup>2</sup>, Glu<sup>4</sup>]deltorphin II. However, the levels of mRNA for the  $\delta$ -opioid receptors in the brain were unaffected (Jenab et al., 1995). Thus, the enhancement of  $\delta$ -opioid receptor agonist induced antinociceptive response by chronic blockade of  $\delta$ -opioid receptors does not appear to be due to changes in either the characteristics ( $B_{\max}$  or  $K_d$ ) or the synthesis (mRNA levels) of the  $\delta$ -opioid receptors. Similarly, chronic i.c.v. administration of [D-Pen<sup>2</sup>, D-Pen<sup>5</sup>]enkephalin resulted in the development of tolerance to its antinociceptive action but the level of brain mRNA for  $\delta$ -opioid receptors was unaffected (Kest et al., 1995).

Naltrindole is a non-selective  $\delta$ -opioid receptor antago-

nist and has been reported to exhibit immunosuppressive activity (Arakawa et al., 1992, 1993). With the development of more selective  $\delta_1$ - and  $\delta_2$ -opioid receptor antagonists, we have recently begun to examine the immunomodulation by selective and non-selective  $\delta$ -opioid receptor agonists and antagonists in vitro (House et al., 1995). The effects of naltrindole as well as of 7-benzylidene-7-dehydronaltrexone and naltriben on the cellular immune function were determined. In vitro exposure to 7-benzylidene-7-dehydronaltrexone resulted in an apparent dose-related suppression of B-cell proliferation, cytokine production by T-helper cells, and natural killer cell activity. Naltrindole was also immunosuppressive for all immune function parameters examined, although it was less active than 7-benzylidene-7-dehydronaltrexone. On the other hand, in vitro exposure to naltriben had no significant effect on any immune function examined (House et al., 1995). These studies clearly indicated that highly selective  $\delta_1$ -opioid receptor antagonist is immunosuppressive whereas  $\delta_2$ -opioid receptor antagonist is not, and provide additional functional evidence for the heterogeneity of  $\delta$ -opioid receptors.

In summary, the present study provides further evidence for the presence of subtypes of  $\delta$ -opioid receptors and further demonstrate that 7-benzylidene-7-dehydronaltrexone and naltriben are highly selective antagonists for the  $\delta_1$ - and  $\delta_2$ -opioid receptors, respectively.

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